

# Finding Correctly Folded Active RNA Motifs

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## abstract

**RNA motifs**, also called active sites, consist of a few functional **modules** separated by unimportant **spacer**.

We have been investigating two well-characterized motifs:



The Isoleucine Aptamer

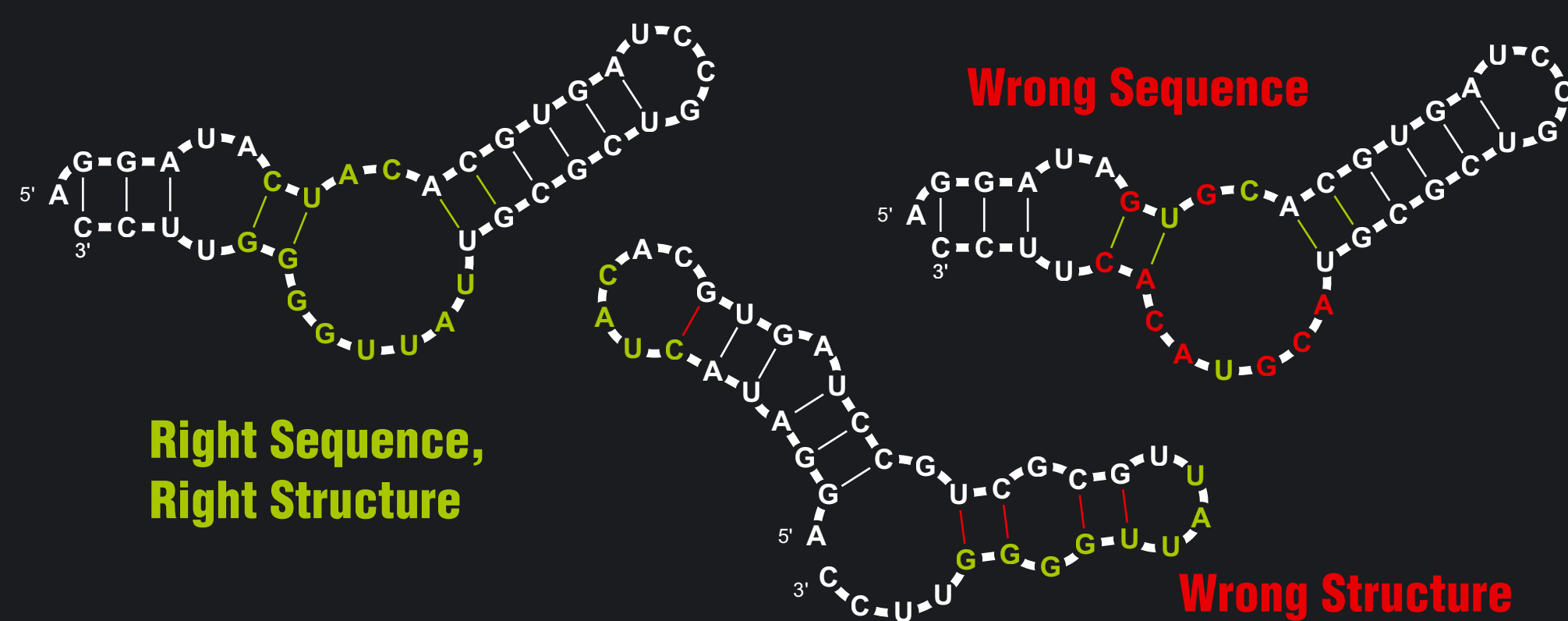
The Hammerhead Ribozyme

Through a combination of **analytical calculations**, **supercomputer simulations**, and **laboratory experiments**, we find that:

- Functional RNA motifs may be far more common than expected, perhaps allowing a **zeptomole RNA world**.
- The **length** and **composition** of randomized RNA pools have dramatic effects on **folding**.

## methods

**Motifs** are defined by inspection of sequence alignments. A motif consists of a set of **modules**. Each module has a characteristic **sequence** and **structure** (where the structure is list of the paired bases). Also, a motif has **rules** specifying helices that must form between its modules.

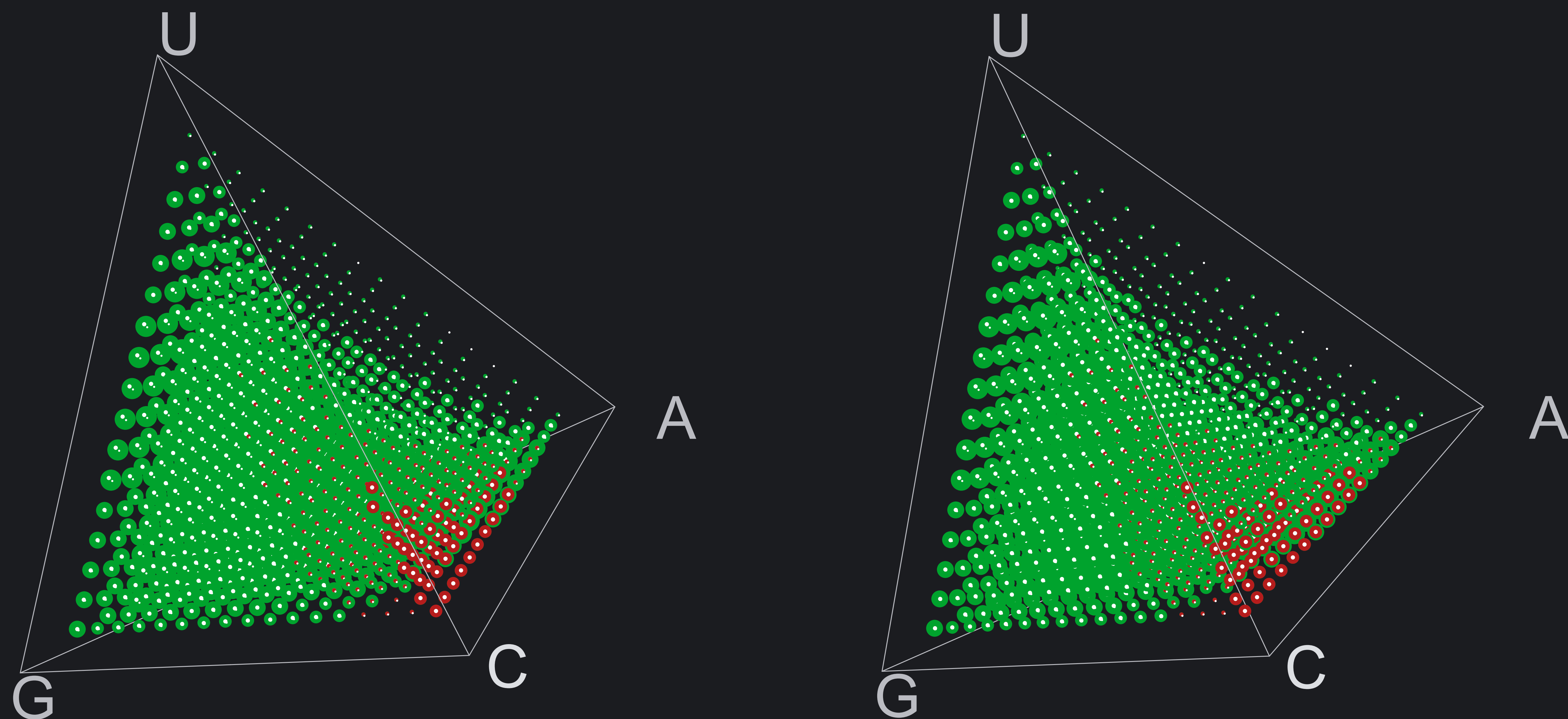


**Frequencies** of motifs in **random sequences** can be calculated analytically, giving  $\Pr(\text{sequence})$ . We must find the probability that at least one of the  $(s+1)!/(s+1-m)!m!$  ways of placing  $m$  modules in  $s$  nucleotides of spacer happens to match all the parts of the motif in order (see calculations section).

**Folding** was characterized using the Vienna RNA package v. 1.4 on the Linux Platinum **supercomputer** at NCSA. We generated sequences for each module, **inserted** them into longer random sequences, and asked how often the **correct base pairs** formed:  $\Pr(\text{structure} | \text{sequence})$ . To get  $\Pr(\text{structure} \& \text{sequence})$ , we multiplied this folding frequency by the frequency of the sequence.

**SELEX** experiments were performed by selecting aptamers against Ile-sepharose, eluting with 10 mM free L-Ile in 50 mM HEPES, 300 mM NaCl, 7.5 mM MgCl<sub>2</sub>, 0.1 mM ZnCl<sub>2</sub>, pH 7.0. Aptamers were selected from pools with 16, 22, 26, 35, 50, and 75 randomized nucleotides over 6-11 rounds of selection, starting with  $2-9 \times 10^{14}$  template DNA molecules. Results were compared with an earlier selection for Ile binders using a similar protocol and a random region of 50 nucleotides.

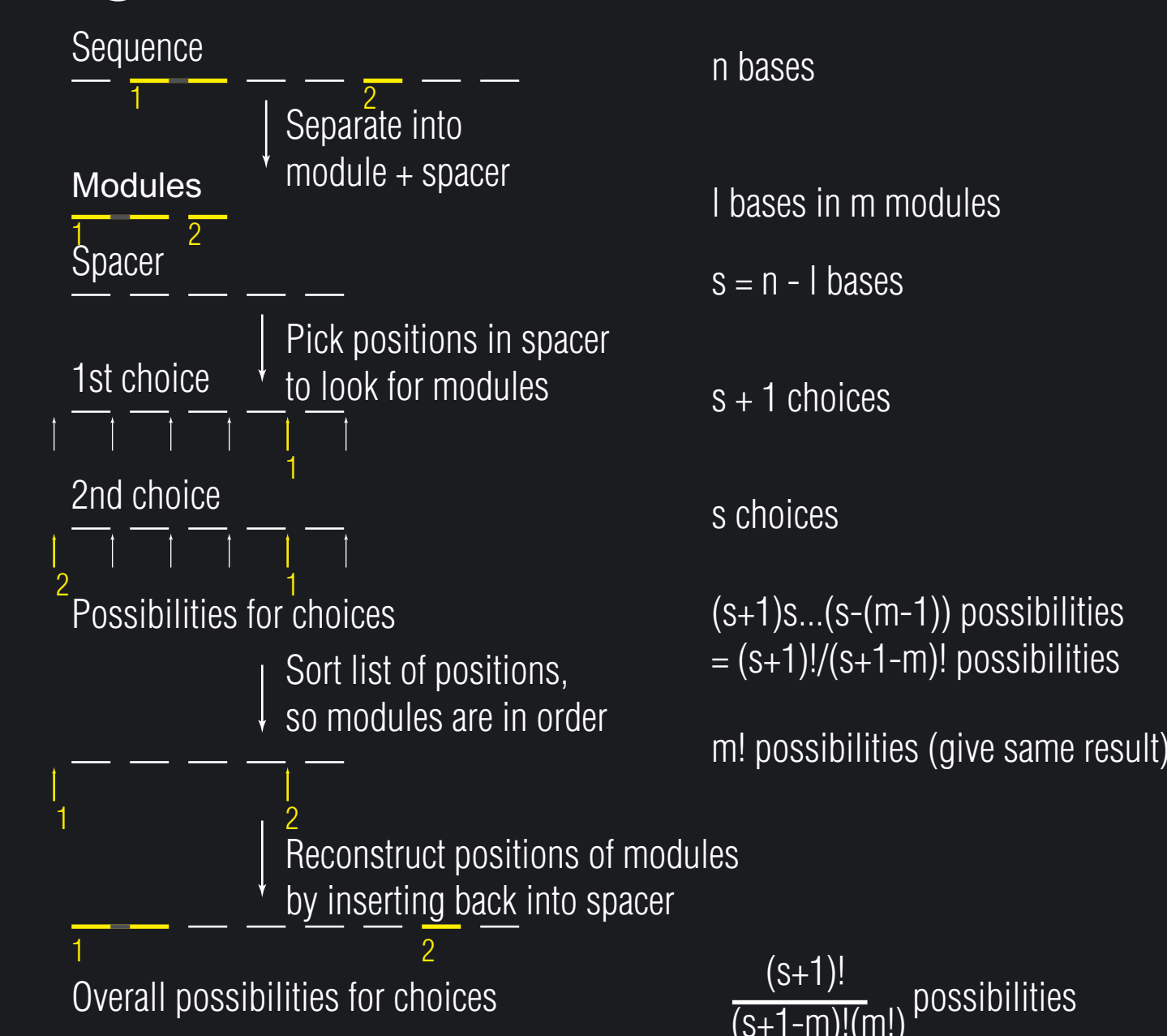
## results



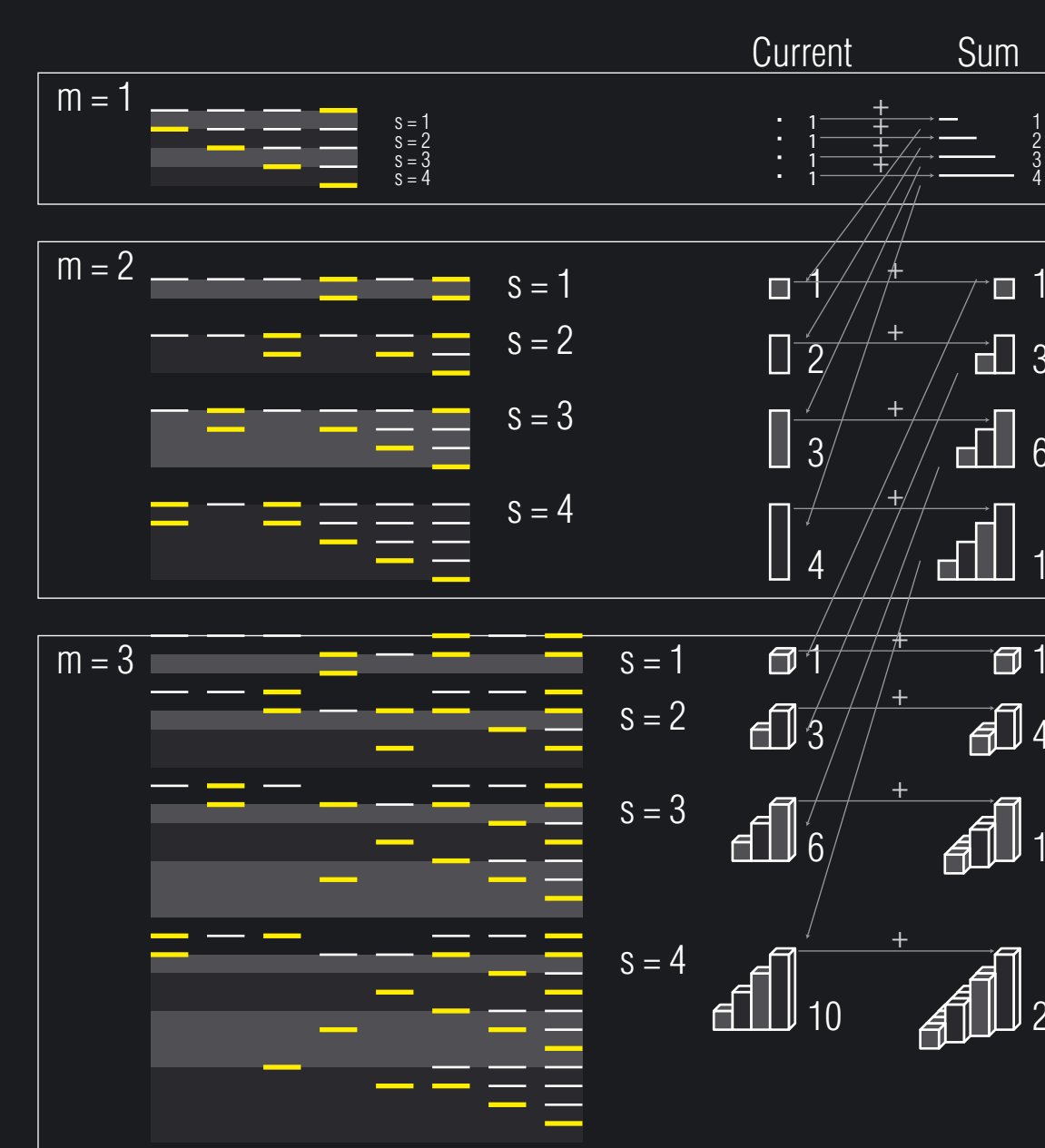
**Stereo pair** showing **folding frequencies** of **Isoleucine aptamers** and **Hammerhead ribozymes** at 5% intervals in the space of possible RNA compositions. Higher-resolution calculations at 2% steps took **600 CPU hours** total spread over **260 processors** on the Linux Platinum cluster at NCSA, stepping over 18,424 compositions with 73,696,000 random sequences (50 nucleotides per sequence). Largest points correspond to 30% correct folding; smallest points to 0.1% correct. **Isoleucine** aptamers fold best in sequences biased towards **purines** (A+C), and typically folds better than the hammerhead site. The **Hammerhead** site folds best with **amino** bases (A+C), probably because of reduced competition.

## calculations

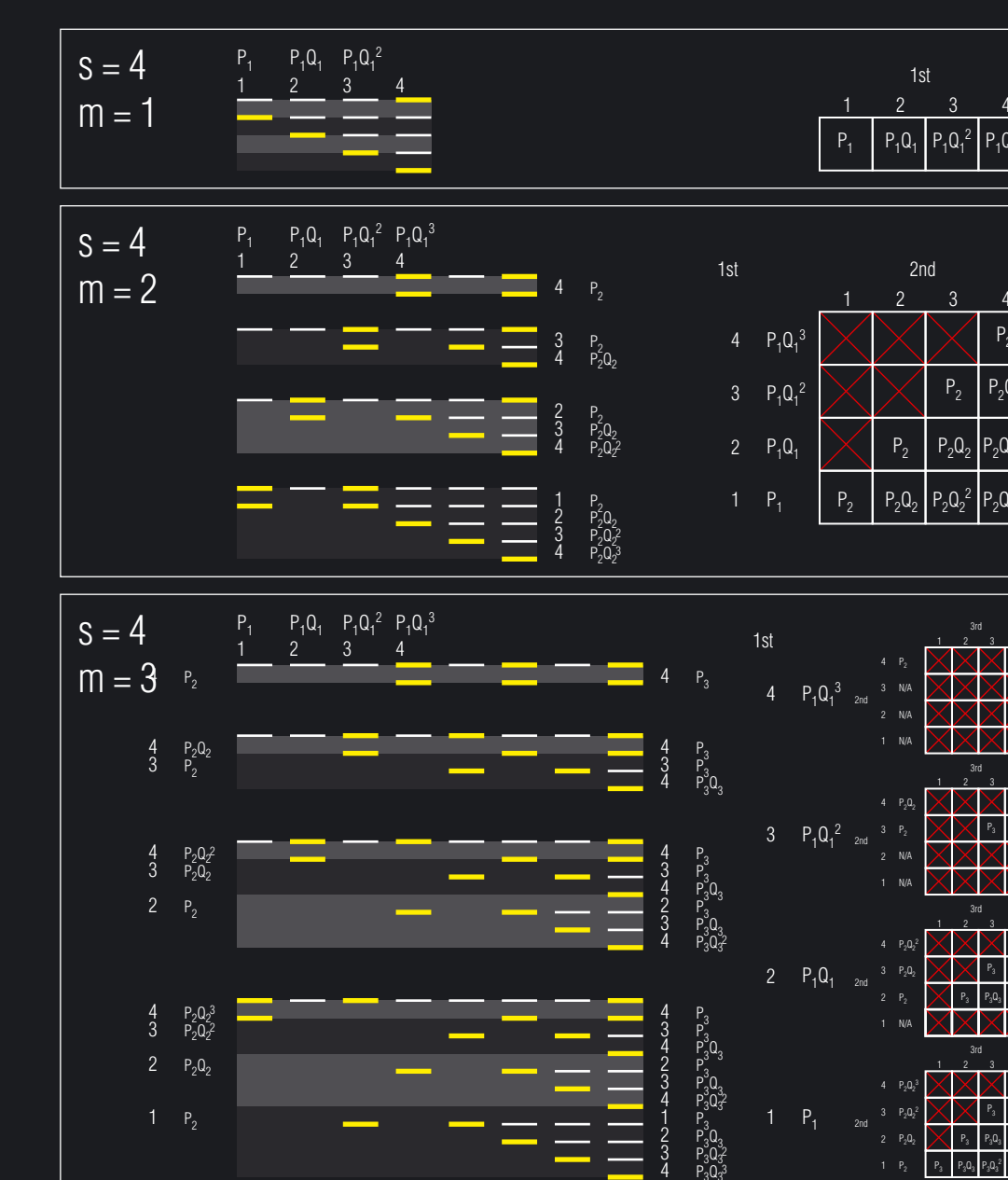
**Finding** several modules in longer sequences is much easier than finding a single conserved region, since there are many ways to place them.



The amount of **spacer s** determines the **size** of the calculation, while the number of **modules m** determines the **dimension**.



Since the same sequence is **resampled**, we calculate the probability **recursively** in terms of smaller problems in lower dimensions.



## conclusions

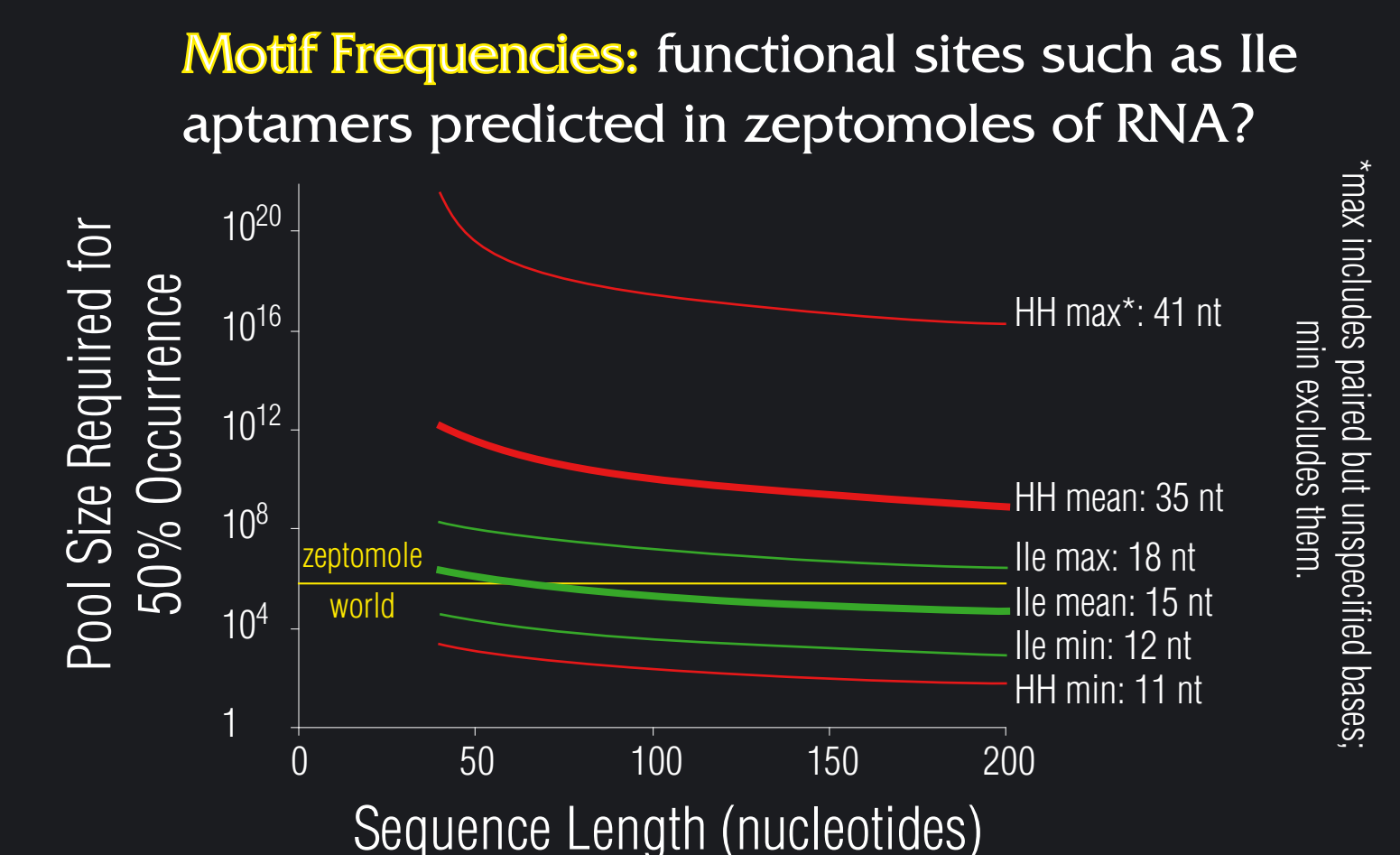
**Modularity** makes functional RNA motifs far more frequent than would otherwise be expected, possibly allowing a **zeptomole RNA world** (functions in less than a million random molecules).

**Selected RNA motifs** should be as short, modular, and evenly divided as possible.

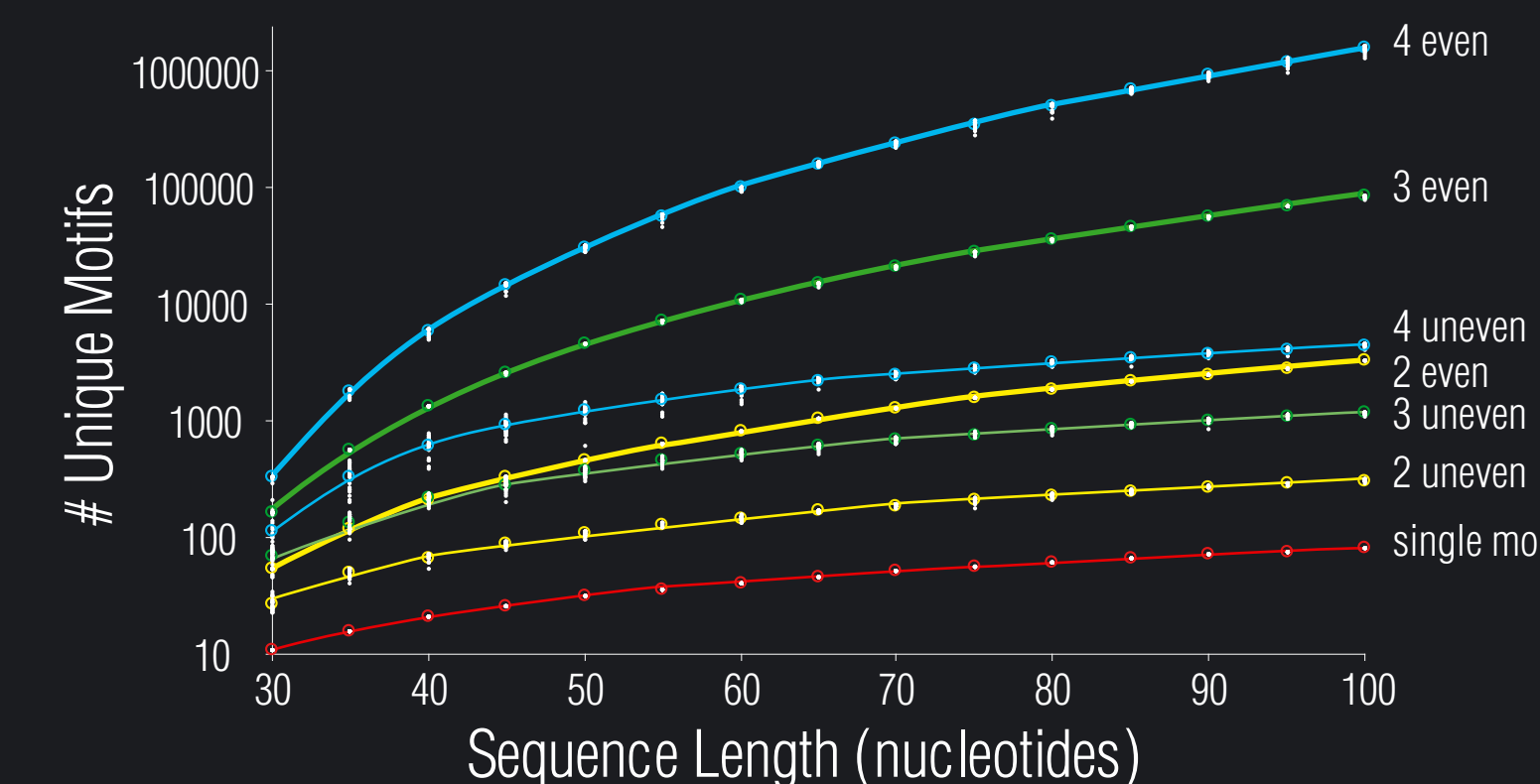
**Primers** have a marked influence on folding; it may be possible to select primer sets that avoid interfering with known active structures.

**Length and composition** of random pools have large effects (several orders of magnitude) on the intrinsic frequencies of functional motifs and on the probability of folding correctly. Length effects on folding in simulations match experiments.

**References:** Knight & Yarus 2003 RNA 9:218-30; Yarus & Knight 2003 in 'The Genetic Code and the Origin of Life', Landes Bioscience; Lozupone et al. in prep; Majerfeld & Yarus 1998 RNA 4:471-8; Schultes et al. 1997 RNA 3:792-806; Sabeti et al. 1997 Chem Biol 4:767-74; Hofacker et al. 1994 Monatsch Chem 125:167-188; Salehi-Ashtiani & Szostak 2001 Nature 414:82-4.



**Calculations** exactly predict the motif sampling in **simulations**: evenness of division critical.



**Length poisoning** reduces the probability of correct folding as the random region increases.

